

Journal of Applied Life Sciences International

20(2): 1-13, 2019; Article no.JALSI.38006

ISSN: 2394-1103

Proximate Nutrient Composition and Antioxidant Properties of *Pleurotus sapidus* 969 Cultivated on *Agave sisalana* Saline Solid Waste

Mutemi Muthangya^{1*}, Mzee J. Amana², Suhaila O. Hashim², Anthony M. Mshandete³ and Amelia K. Kivaisi³

¹Department of Biochemistry, South Eastern Kenya University, P.O.Box 170 – 90200, Kitui, Kenya.

²Department of Biochemistry, Pwani University, P.O.Box 195 - 80108, Kilifi, Kenya.

³Department of Molecular Biology and Biotechnology, College of Natural and Applied Sciences,

University of Dar es Salaam, P.O.Box 35179, Dar es Salaam, Tanzania.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JALSI/2019/v20i230081

Editor(s

(1) Dr. Kuldip Singh, Department of Biochemistry, Govt. Medical College, Punjab, India.

Reviewers:

(1) Fatih Kalyoncu, Manisa Celal Bayar University, Turkey.

(2) Hasan Hüseyin Doğan, Biology, Selcuk University, Turkey. (3) Dr. Salisu Muhammad Tahir, Kaduna State University, Kaduna, Nigeria.

(4) Dr. Ndomou Mathieu, Department of Biochemistry, University of Douala, Cameroon.

Complete Peer review History: http://www.sdiarticle3.com/review-history/38006

Original Research Article

Received 25 October 2017 Accepted 11 January 2018 Published 04 April 2019

ABSTRACT

Effects of pure and mixed substrates of sisal waste, grass (*Panicum coloratum*) and a combination of the two substrates at 50:50 (w/w) on nutritional composition, minerals and antioxidant potential of sun-dried *Pleurotus sapidus* 969 were investigated in the present study. To determine the proximate chemical composition and antioxidant properties of the samples, standard analytical procedures were employed. Moisture content, crude protein and crude fibre ranged between 11.09-12.80%, 6.4-6.6% and 18.3-30.5%, respectively. Macro elements Ca, Mg, Na, K, and P were also found in substantial amounts with K being present in an exceedingly higher amount (541.3-657.1 mg/100g) than the other macro minerals. The samples from the three substrates contained antioxidant β -carotene (4.6-6.0 mg/100g), lycopene (4.9-5.1mg/100g), Vitamin C (5.2-5.6 mg/100g), phenols (361.0-859.0 mg of GA/g) and flavanoids (33.5-64.0 mg RE/g).Mushroom harvested from

mixed substrates contained better nutritional qualities than the pure substrate, although the phenolic content in mushrooms cultivated on sisal substrate was higher. The results further showed that, all the extracts exhibited scavenging ability and metal chelating activity. The findings showed that *Pleurotus sapidus* 969 is rich in nutrients, macro minerals as well as natural antioxidant which could be explored for pharmaceutical applications.

Keywords: Sisal; antioxidant; free radicals; Pleurotus; flavanoids; phenols.

1. INTRODUCTION

Cultivation of the oyster mushroom, Pleurotus spp has increased greatly throughout the world during the last few decades and constitute the second largest variety of mushrooms produced in the world [1] with China being the primary source. Pleurotus cultivation has the advantage of being cultivable in tropical climates, simple to produce, and compatible with organic substrates rich in lignin and cellulose. Their ability to utilize different substrates has made them the subject of broad research that generally mentions their nutritional quality and the effect of substrate variation on the primary metabolites that are directly related to the nutritional quality. Mushrooms have greatly varied and important uses throughout the world [2]. Mushrooms are valuable health foods since they are poor in calories, fat, and essential fatty acids, and rich in proteins, vitamin and minerals [3]. Moreover, their medicinal properties have been reported such as anti-tumour and immunomodulating effects [4], reduction of blood cholesterol concentrations, prevention or alleviation of heart disease and reduction of blood glucose levels [5]. These properties of mushrooms have been reported by Ferreira et al. [6], do be as a result of the bioactive products with antioxidant potential (sterols, tocopherols, flavonoids, Carotenoids and phenolic compounds [6].

Sequences of chemical reactions result in an imbalance between oxidant and antioxidant reactions and are typically referred to as oxidative stress [7]. Both classes of substances (oxidants and antioxidants) are generated in an oxidation-reduction (redox) set-up, [8] and has been implicated as causes of degenerative diseases such as atherosclerosis, cancer, and tissue damage in rheumatoid arthritis [9] Reactive species are commonly identified as substances leading to the oxidation of lipids (epoxidation), glucose (glycation) and proteins (carbonylation). Maintenance of equilibrium between free radicals production and antioxidant defences is an essential condition for normal organism functioning [10]. Non-controlled production of free radicals has been attributed to various kinds of cancer and diabetes according to Ferreira et al. [6]. Natural products with antioxidant activity, in particular mushrooms, are used to aid the endogenous protective system, increasing interest in the antioxidative role of functional foods or nutraceutical products [3]. Antioxidants pay an important role in the prevention and treatment of a variety of diseases by removing free radical intermediates and inhibit other oxidation reactions by being oxidized themselves [11]. Many studies have found that some species of mushrooms are having therapeutic properties [12] due to a wide variety of free radicals or reactive oxygen species scavengers which have made them attractive as nutritionally beneficial foods and as a source for drugs development [13]. According to Barros et al. [14], mushroom flavonoids can act as free radical scavengers to terminate the radical chain reactions that occur during the oxidation of triglycerides in the food system. Apart from being a delicacy and tasty foods, mushrooms have been reported to have special biochemical compositions, with significant contents of antioxidant compounds, proteins, minerals, vitamins and water, which attract more attention as functional health promoters [15].

The chemical composition and nutritional quantity of edible mushrooms have been reported previously [16]. Studies have consistently shown an inverse association between consumption of vegetables and fruits and the risk of certain forms of cancer [17]. However, the protective effects have been primarily attributed to well-known antioxidants. such as ascorbic acid and other related compounds [18]. Different mushrooms species have been studied for new therapeutic alternatives and the results proved their bioactive properties [19]. Mushrooms are rich sources of nutraceuticals [20], which are responsible for antioxidant content [21]. Recent investigations revealed that polysaccharides and extracts of mushrooms had strong antioxidant and no synthase activation properties [22,23,24]. According to Muhammad Nasir et al. [25], there

are about 5000 different species of mushrooms, of which at least 1220 are reported to be edible. There are about 40 species under *Pleurotus* mushroom, in that 25 species are commercially cultivated [26]. Most of these cultivated mushrooms are consumed as food or food ingredients in various food preparation and processed food products. This has led to the growing interest in the use of edible mushrooms extracts as dietary supplements based on the facts that they have a lot of bioactive compounds.

Pleurotus mushrooms can be grown on various agro-residues (as substrate) as reported by Muthangya et al. [27]. The mushroom cultivation substrate has been reported to influence its growth, yield as well as the functional, organoleptic and chemical [28]. This study was therefore designed to investigate the nutritive and antioxidant property of P. sapidus 969 cultivated on Agave Sisalana saline solid waste and on grass (Panicumcoloratum) as well as on a mixture of the two substrates at 50:50 (w/w) as reported in Muthangya et al. [29].

2. METHODS

2.1 Samples of *Pleurotus sapidus* 969 Mushrooms

P. sapidus 969 mushrooms used in this study were cultivated on pre-treated saline sisal leaf decortications waste as reported in Muthangya et al. [29]. Mushrooms were sundried on a fabricated solar drier for 7 hours on a full sunny day before analysis.

2.2 Determination of Moisture, Crude Fibre and Macro Element Content

The sun-dried *P. sapidus* 969 mushrooms were analysed for moisture and total fibre content using a Near Infrared Reflectance Spectroscopy (NIRS). The NIRS technique uses near-infrared light, instead of chemicals as in conventional "wet chemistry" methods. The samples were prepared and analysed as described by Windham et al. [30]. The prepared mushrooms samples were analysed for Ca, Mg, Na, K, and P, according to AOAC (2000).

2.3 Crude Protein Determination

Crude protein in *P. sapidus* 969was determined according to the method previously reported by Tibuhwa et al. [31]. A known weight of each

mushroom sample was taken and digested using the micro Kjeldahl method. After completion of digestion, organic nitrogen was determined calorimetrically using the Indophenol-blue method and $\mathrm{NH_4}^+\text{-N}$ as standard. The absorbance was measured at 660 nm. The total crude protein was obtained and calculated as described in Allen (1989).

2.4 Mushroom Crude Extracts Preparation

The mushroom crude extract was prepared in ethanol according to Tibuhwa, [32], with modification, where 1gm of dried whole mushrooms fruiting body was weighed at room temperature (29±3°C). The samples were finely crushed using motor and pestle, and extracted with 250 ml of ethanol as a solvent. The crushed powder was constantly stirred for 48 hrs and thereafter filtered using Whatman number 4, filter paper. The filtrates were evaporated to dryness in a rotary evaporator 90 rpm under reduced pressure and at 40°C. The concentrated extracts obtained were stored in the dark at 4°C until further analysis. The yields of evaporated dried extracts were obtained by the gravimetric method. The percentage yield of the extracts was calculated based on dry weight as:

Yield (%) =
$$\frac{(W_1 \times 100)}{W_2}$$

Where:

 W_1 = weight of extract after ethanol evaporation W_2 = Weight of the ground mushroom powder

2.5 Quantitative Antioxidant Assay

2.5.1 Determination of total phenolics content (GAE/g)

The concentration of phenolic compounds in the extract of *P. sapidus* 969 mushroom was measured by Folin-Ciocalteu colourimetric method according to the method previously reported by Tibuhwa [32], with modification. A blue colour was developed by reaction of phenolic compounds and Folin-Ciocalteu's reagent. The extract solution (1 ml) was mixed with 1 ml of Folin-Ciocalteau reagent and after 3 min, 0.8 ml of 7.5% (w/v) sodium carbonate was added to the mixture. The reaction was kept in the dark for 30 min with agitation and thereafter centrifuged at 3300 g for 5 min. The absorbance was measured at 765 nm and total phenolic

content was expressed as gallic acid equivalent (GAE) to 1 g per extract using gallic acid as a standard.

2.5.2 Determination of total flavonoid

Determination of total flavanoids was carried out using the aluminium chloride colourimetric method according to Jaita et al. [33], as reported in Tibuhwa [32]. Each extract (1 ml) was diluted with 4.3 ml of 80% aqueous ethanol containing 0.1 ml of 10% aluminium nitrate and 0.1 ml of 1M aqueous potassium acetate. The mixture was incubated for 40 minutes at room temperature and the absorbance determined colourimetrically at 415 nm. A standard curve of flavonoids was prepared and concentration of flavonoids in the test samples determined.

2.5.3 β -carotene and Lycopene contents

 β -carotene and lycopene were determined according to the method of Nagata and Yamashita, [34]. In brief, 100 ml of mushroom extract (10 mg/ml) was vigorously shaken with 10 ml of acetone-hexane mixture (92:3) for 1 min. and filtered through Whatman number 4 filter paper. The absorbance of the filtrate was measured at 453, 505 and 663 nm. β -carotene and lycopene contents were calculated according to the following equations:

Lycopene (mg/100mg) =
$$0.0458 A_{663} + 0.372 A_{505} - 0.0806 A_{453}$$

$$\beta$$
-carotene (mg/100mg) = 0.216 A₆₆₃ - 0.304 A₅₀₅ + 0.452 A₄₅₃

2.5.4 Determination of vitamin C

The vitamin C content was determined diametrically using 2, 6 DichlorophenoIndophenoI methods according to Plumer [35]. One (1) gram of grounded sample was mixed with 25 ml of 5% metaphosphoric acid solution and shaken for 30 min. The mixture was then filtered through Whatman no. 42 filter paper using suction pump. Ten (10) ml of the filtrate was titrated against 0.025% of 2.6 DichlorophenoI IndophenoI reagents. The amount of vitamin C in each extract was calculated from the equation:

Ascorbic acid mg/100 g =
$$\frac{A \times I \times V \times 100}{V_2 \times W}$$

Whereas

A = Quantity of ascorbic acid (mg) reacting with 1ml of 2, 6 Indophenol

I = Volume of indophenol (ml) required for the completion of extract titration

 V_2 = Total volume of extract

W = Weight of the ground mushroom

2.6 DPPH Free Radical Scavenging Activity

The scavenging ability on 1,1-diphenyl-2-picrylhydrazyl (DPPH) radicals was determined according to the method of Masuda et al. [36], and Jaita et al. [33], as previously reported by Tibuhwa et al. [31]. Each extract (0.01-0.14 mg/ml) was mixed with 1 ml of methanolic solution containing DPPH radicals (0.4 mM). The mixture was shaken vigorously and left to stand for 30 min in the dark. The absorbance was measured at 515 nm. The percentage of DPPH radical scavenging activity of each extract was determined within the range of dose-response and was calculated as:

DPPH radical scavenging activity (%) =

$$\frac{A_0 - (A_1 - A_s) * 100}{A_0}$$

Where

A₀ = Absorbance of the control solution containing only DPPH

A₁ = Absorbance in the presence of mushroom extract in DPPH solution

A_s = The absorbance of the sample extract solution without DPPH

The EC50 value (total antioxidant necessary to decrease the initial DPPH radical concentration by 50%) was determined from a plot of scavenging activity against the concentration of extracts.

2.7 Chelating Effect on Ferrous Ions

The ability of *P. sapidus* 969 extracts to chelate ferrous ions was estimated by the method of Dinis et al. [37]. The extract (1 mg/ml) was added to a solution of 2 mM ferrous chloride (0.05 ml). The reaction was initiated by the addition of 5 ferrozine (0.2 ml) and the mixture was then shaken vigorously and left to stand at room temperature (28-30°C) for 10 min. The absorbance of the solution was measured spectrophotometrically at 562 nm. The percentage inhibition of ferrozine-Fe²⁺ complex formation was calculated as;

$$\{(A_0 - A_1) / A_0\} \times 100$$

Where

 A_0 = absorbance of the control

A₁ = absorbance in the presence of the mushroom extract

2.8 Statistical Analysis

The experimental results were expressed as mean \pm SD (Standard deviation) of n=3 measurements. Statistical analysis of the data was carried out using student's t-test and the results were considered significant when P= .05.

3. RESULTS AND DISCUSSION

3.1 Composition of Sun-dried *Pleurotus* sapidus 969

The moisture contents of the dried *Pleurotus* sapidus 969were found 11.9-12.8% (Table 1) with no significant difference at *P*= .05 level. The highest moisture content was found in *P.sapidus* 969cultivated on a mixture of sisal and grass substrate (1:1), followed by grass and the least was in sisal alone.

The fibre content was found highest in $P.\ said$ 969 (6.6g/100g) cultivate on grass. The variation in fibre content between the mushrooms from the three different substrates was not statistically significant at P=.05. Comparison of the results of the protein content of the mushroom from the three substrates showed a significant difference at P=.05 with the highest crude protein content being recorded from the mushrooms in the combined substrate of sisal and grass (30.5%).

The results of obtained by this study for the dried *Pleurotus sapidus* 969 are within the range of those reported previously for other *Pleurotus* species. Muthangya et al. [27], working on *Pleurotus* HK 37 from the same substrate reported results, which were within the range of those obtained in this study. Oyetayo and Ariyo, [38], working on *Pleurotus ostreatus* reported the moisture content of dried samples to be within 9.00-10.72%. While previously, Chang and Miles [39], reported the moisture content of dried mushrooms to be in the range 9 - 13%. Sales-

Campos et al. [40], reported a variation in fibre content while working on several Pleurotus sp. grown on crushed sugar cane, elephant grass and banana tree leaves, on the other hand, the results obtained on the fibre content was within the range (5.4–30.0%) previously reported by other authors for Pleurotus sp. [41] cultivated on different substrates. The protein contents of mushrooms are reported to vary according to genetic structure of species, physical and chemical differences in growing [42], cultivation time and strain [43,44], as well as the stage of development and level of nitrogen available [39]. The mushroom protein contents that were found in this study (Table 1) are in agreement with the range of mushroom protein contents reported in the literature [43]. Varying between 17 and 42.5%, but higher in *P.sapidus* 969cultivated on grass and on a combined substrate of grass and sisal than the value (20.28%) reported by Bonatti et al. [45] for Pleurotus ostreatuscultivated on cotton waste. The present results showed that protein content of P. sapidus 969 was significantly higher when the mushroom was cultivated on a combination of sisal and grass than that obtained for the mushroom grown on separate substrates.

3.2 Macro-minerals Elements

Pleurotus sapidus 969 mushroom samples analysed in this study contained macro-minerals including; calcium, magnesium, sodium, potassium and phosphorus (Table 2).

The highest amount of Ca (7.7 mg/100 g) was recorded in the *P. sapidus* 969 samples from grass substrate, followed by sisal: grass (7.6 mg/100 g) and lastly sisal (6.1 mg/100 g). Mg concentration was the highest in sisal: grass samples (18.1 mg/100 g) and the least in samples obtained from sisal substrate (16.21 mg/100 g). The value of Na, K and P in the *P. sapidus* 969 were found to be in the range of 14.2-16.46, 541.2-657.1 and 123.4-131.7 mg per 100 g, respectively. Minerals in human diets are essential constituents for metabolic reactions, transmission of nerve impulses, healthy bone formation, regulation of water and salt balance

Table 1. Composition (%)of sun-dried *Pleurotus sapidus* 969 and crude extract yields, Mean±SD, n=3)

Cultivation substrate	Moisture (%)	Total fibres (%)	Crude proteins (%)	Crude extract yield (%)
Sisal	11.9±0.03	6.4±0.1	18.3±0.4	17.0±0.4
Grass	12.2±0.01	6.6±0.2	23.3±0.2	17.9±0.3
Sisal: Grass	12.8±0.04	6.5±0.2	30.5±1.2	13.7±0.4

Table 2. Macro-minerals composition of P. sapidus 969(g/100g of dried sample) Mean±SD, n=3

Cultivation substrate	Macro-minerals (mg/100 g)					
	Са	Mg	Na	K	Р	
Sisal	6.1±0.4	16.21±0.6	15.17±0.1	614.5±1.9	117.7±0.9	
Grass	7.7±0.3	17.8±0.5	14.2±0.3	541.3±2.2	123.4±0.9	
Sisal : Grass	7.6±0.1	18.1±0.4	16.4±0.2	657.1±4.8	131.7±2.0	

Kalac, and Svoboda [46]. The mineral contents of P. sapidus 969from the two different substrates and their combinations in this study did not vary significantly at P=.05. The results of the macro-minerals elements composition of P.sapidus 969 are within the range as those reported by Muthangya et al. [27], from dried samples of Pleurotus HK 37 cultivated on the same substrates, although slightly higher. The values of calcium in this study are an indication P. sapidus 969 is a valuable food for formation and maintenance of bone and normal function of nerves and muscles in humans and other vertebrates as reported by Wang et al. (2010). Mg, an essential co-factors for certain enzymes in various biochemical pathways was detected in P. sapidus 969and the levels of Mg were quite higher than those reported (1.69-3.57 mg/100 g) for Pleurotus ostreatuscultivated on different woody substrates [38]. Na and K are important in the maintenance of osmotic balance between cells and the interstitial fluid in animal systems [47]. These results indicate that these mushrooms could play a role in human health by lowering blood pressure, reducing the risk of osteoporosis and in maintaining bone health (Wang et al., 2010). The results of phosphorus in this study (123.4-131.7 mg/100 g) compare well with 122.28 mg/100 g reported for a wild P. ostreatus [47]. The differences in phosphorus contents in mushroom have been attributed previously to substrates what about mushroom species/strain since they differ in substrate utilization/absorption and translocation biomaterials from substratesused for growing the mushrooms according to Ahmed, [48]. Pleurotus species canprovide a useful source of phosphorus. potassium. calcium. magnesium. Thus, the inclusion of P. sapidus 969 in the diet could be oneof the strategies for combating macronutrient deficiencies

3.3 Antioxidant Contents of *Pleurotus* sapidus 969

3.3.1 Total phenol and flavonoid contents

The total phenolic and flavonoid content in Pleurotus sapidus 969analysed in this study are shown in Fig. 1. The total phenolic and flavanoids contents in the mushroom samples were 859.0, 784.7 and 361.0 mg of GA/ g and 33.5, 64.0 and 53.8 mg RE/g in the mushrooms grown on sisal, grass and sisal: grass substrates, respectively.

The findings of this study is supported by previous findings of Phenolic compounds in mushrooms as reported by Tibuhwa, [32] and linked to various biological functions including antioxidant activity Phenolic compounds are well known secondary metabolites commonly found in plants and mushrooms and reported to have vital biological functions including antioxidant activity [49]. Knowing the amount of total phenolic compounds in mushrooms is of great importance in their nutritional and functional characterization since the profile of the phenolics has been reported to be species-specific Banerjee et al. [50]. Phenolic compounds have been reported to be of great interest due to their possible use as dietary supplements or food preservatives, Jayakumar et al. [51]. Several species of mushroom have been reported to contain a wide variety of free radicals or reactive oxygen species scavengers, which have made mushrooms attractive as nutritionally beneficial foods and as a source for drugs development [13]. Barros et al. [14] reported that mushroom flavonoids can act as free radical scavengers to terminate the radical chain reactions that occur during the oxidation of triglycerides in the food system. Flavonoids have been reported to decrease capillary fragility and exert a cortisonelike effect on tissues [52] and protect against cancer and heart diseases [53]. It, therefore, implies that the high flavonoids content in the mushroom extracts might be responsible for the therapeutic effect of some mushroom species earlier [54].

Previous studies have shown that food consumption with high phenolic content can reduce the risk of heart disease [55]. From this study, the high levels of phenols and flavanoids make *P. sapidus* 969 favourable for nutritional and therapeutic application as supported by the findings of Ferreira et al. [56].

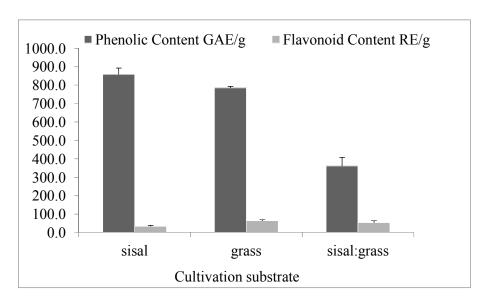


Fig. 1. Total phenol and flavonoid contents of *P. sapidus* 969, Values are expressed as mean ± SD mg of Gallic acid equivalent per gram of dry weight (mg GAE/gm)

3.3.2 β-carotene, lycopene and vitamin C content

Carotenoids are natural colourants, stabilizers and active in the protection process of human body cells, where they balance and offset the destructive effects of free radicals Jayakumar et al. [51]. The quantities of β -carotene, Lycopene and Vitamin C content of P. sapidus 969analysed in this study are presented in Fig. 2. The content of β -carotene was in the range of 4.6 mg/100g to 6.0 mg/100g, lycopene was in the range of 4.9 mg/100g to 5.1 mg/100g, while vitamin C was in the range of 5.2 mg/100g to 5.6 mg/100g in the are major substrates. Carotenoids antioxidants with known health benefits, while diets high in lycopene; a cyclic isomer of β carotene has been linked to reduction of prostate cancer and cardiovascular diseases [57], whereas. Ascorbic acid is reported to directly interact with radicals in plasma, preventing damage to red cell membranes [51].

The results of β -carotene, lycopene and vitamin C obtained in this study are within the range of those reported previously by Muthangya et al. [27] from sun-dried samples of *Pleurotus* HK 37 cultivated on similar substrates. The presence of these compounds in *P. said* 969 is an indication that these mushrooms are equipped with antioxidant properties. Jayakumar et al. [51] reported similar findings of carotenoid and ascorbic acid compounds from *P. ostreatus*mycelium extracts. The quantities of

these compounds in various extracts have been suggested to be influenced by the culture medium used for producing the [58], a similar scenario observed in this study where different substrates were used to cultivate *P. said* 969. These findings support Barros et al. [59], who reported that the carbon source and especially the nitrogen sources have a direct influence on the quantum of biologically active substances in the extracts.

3.4 Antioxidant Activities

3.4.1 DPPH Free radical scavenging activities

The result from this study showed that the free radical scavenging activity of *P. sapidus* 969extract from the three cultivation substrates increased with increasing concentration of extract indicating the concentration dose dependency of anti-oxidative activities (Fig. 3). This observation concurs with that of Banerjee et al. [50] who also noted a similar trend of anti-oxidative activities dose dependency and associated it with the presence of reductones that are reported to be the terminators of free radical chain reactions.

In this study, the maximum scavenging activity values were at a dilution of 0.14mg/ml. The mushroom extracts from grass substrate showed the highest percentage (96.7%) scavenging power while the extracts from sisal and sisal: grass had 84.7% and 71.2%, respectively.

However, the total antioxidant necessary to decrease the initial DPPH radical concentration by 50%, determined from plotted graph of scavenging activityagainst different concentration of the extracts, showed the extract from sisal had the highest ability (EC50 < 0.09 mg/ml) followed by that from grass (EC50 < 0.11 mg/ml) while the extracts from sisal:grass had the least ability of (EC50 < 0.13 mg/ml), a similar observation reported by Muthangya et al. [27], working dried samples of Pleurotus HK 37 cultivated on the same substrates. This result shows that the P. sapidus 969mushroom studied have high ability scavenging compared to other mushrooms. Although in this study, mushrooms from sisal: grass had the least ability of (EC50 < 0.13 mg/ml), this value is still better compared to other well appreciated antiradical mushrooms. Filipa al., [60] established EC50 values in Paxillus involutus and Pisolithusarhizus of (EC50) = 0.61 and EC50 = 0.56 mg/ml), respectively which show them having relatively low free ability radical scavenging compared mushrooms from sisal: grass with least ability in this study. The higher content of phenolic compounds in mushrooms cultivated on sisal substrate could be the cause of the high total antioxidant necessary to decrease the initial

DPPH radical concentration by 50% an observation in line with the findings of Abdullah et al. [61] working on of Brazilian button mushrooms.

3.4.2 Chelating ability of ferrous ions

Fig. 3 depicts the iron chelating ability of *Pleurotus sapidus* 969 cultivated onthe different substrates under investigation in this study. The ferrous ion-chelating effect of all samples increased well with increasing concentrations (Fig. 4). *P. sapidus* 969 from sisal substrate had the highest iron chelating ability (90.8% at 0.12 mg/ml), while the weakest metal chelating ability (82.2%) was recorded for samples from a combined substrate of sisal and grass.

Extract from samples cultivated on sisal substrates recorded 86.3% metal chelating ability at the same concentration. It has been observed that metal ion chelating antioxidants would also remove the oxidative damage from other less prominent but equally damaging pro-oxidant metal ions such as Cu [62]. Thus, the iron chelating capacity of the mushroom species would prevent transition metals to participate in the initiation of oxidative stress.

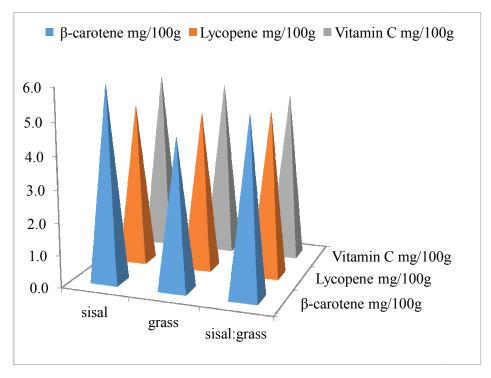


Fig. 2. Total β -carotene, Lycopene and Vitamin C, the content of *P. sapidus* 969 Values are expressed as mean \pm SD mg/100g

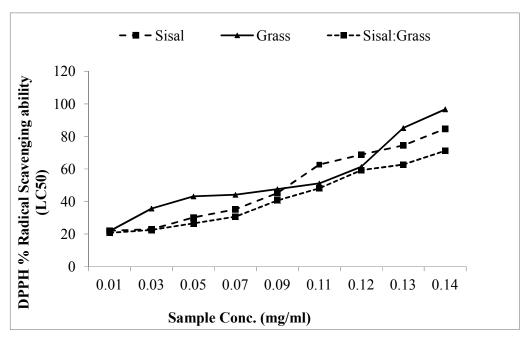


Fig. 3. DPPH radical scavenging activity (%) of *P. said* 969 (ethanolic extract) cultivated on sisal grass, sisal: grass at 1:1 Values recorded are (mean ± SD, n=3)

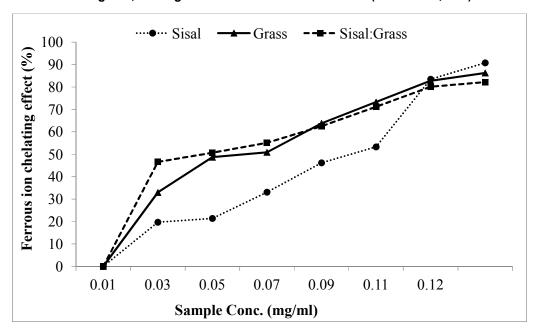


Fig. 4. Ferrous ion chelating effect (%) of *P. said* 969 (ethanolic extract) cultivated on sisal grass, sisal: grass at 1:1

4. CONCLUSION

It was observed that fruiting bodies harvested from different substrates varied in their biochemical analysis. It might be due to the variability of the substrates to provide different

nutritional elements to mushroom grown on these substrates. Among the substrates investigated in this study, a combination of Sisal and grass gave the best overall composition of all the nutrients. The nutritional and antioxidant investigations on the mushroom cultivated on the different substrate revealed that all the mushrooms possess high reductive potential and metal chelation activities, with high concentration of macro nutrients, proteins, total phenol and total flavonoids. These bioactive compounds together with the high antioxidant activities of *P. sapidus* 969 could be explored as a natural rich antioxidant food, which may enhance the immune system against oxidative damage, or it may be utilized as a potential source for drug development.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Mshandete AM. Cultivation of Pleurotus HK-37 and Pleurotus sapidus (oyster mushrooms) on cattail weed (*Typha* domingensis) substrate in Tanzania. International Journal of Research in Biological Sciences. 2011;1(3):35-44.
- Wan Rosli WI, Solihah MA, Mohsin SSJ.
 On the ability of oyster mushroom
 (*Pleurotus sajor-caju*) conferring changes
 in proximate composition and sensory
 evaluation of chicken patty International
 Food Research Journal. 2011;18(4):1463 1469.
- Reis FS, Barros L, Martins A, Ferreira ICFR. Chemical composition and nutritional value of the most widely appreciated cultivated mushrooms: an interspecies comparative study. Food Chem. Toxicol. 2012;50:191-197.
- Ferreira ICFR, Vaz JA, Vasconcelos MA, Martins A. Compounds from wild mushrooms with antitumor potential. Anti Canc. Agents Med. Chem. 2010;10:424-436.
- Jeong SC, Jeong YT, Yang BK, Islam R, Koyyalamudi SR, Pang G, Cho KY, Song CH. White button mushroom (Agaricus bisporus) lowers blood glucose and cholesterol levels in diabetic and hypercholesterolemic rats. Nutr. Res. 2010;30:49-56.
- Ferreira ICFR, Barros L, Abreu RMV. Antioxidants in wild mushrooms. Curr. Med. Chem. 2009;16:1543-1560.
- Poli G, Schaur RJ, Siems WG, Leonarduzzi G. 4-Hydroxynonenal: A membrane lipid oxidation product of

- medicinal interest. Medicinal Research Reviews. 2008:28:569-631.
- 8. Ralser M, Wamelink MM, Kowald A, Gerisch B, Heeren G, Struys EA, Klipp E, Jakobs C, Breitenbach M, Lehrach H, Krobitsch S. Dynamic Rerouting of the Carbohydrate Flux Is Key to Counteracting Oxidative Stress. Journal of Biology. 2007;6:10.
- Jang HD, Chang KS, Huang YS, Hsu CL, Lee SH, Su MS. Principal phenolic phytochemicals and antioxidant activities of three Chinese medicinal plants. Food Chemistry. 2007;103:749–756.
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int. J. Biochem. Cell Biol. 2007;39:44-84.
- Sies H. Oxidative stress: Oxidants and antioxidants. Experimen Physiol. 1997; 82(2):291-95.
- 12. Oyetayo VO. Free radical scavenging and antimicrobial properties of extracts of wild mushrooms. Brazilian Journal of Microbiology. 2009;40(2):380-386.
- Guerra-Dore CMP, Azevedo TCG, de Souza MCR, Rego LA, de Dantas JCM, Silva FRF, Rocha HAO, Baseia IG, Leite E L. Antiinflammatory, antioxidant and cytotoxic actions of β-glucanrich extract from Geastrum saccatum mushroom. Int Immunopharmacol. 2007;7(9):1160-69.
- Barros L, Cruz T, Baptista P, Estevinho L M, Ferreira ICFR. Wild and commercial mushrooms as a source of nutrients and nutraceuticals. Food Chem Toxicol. 2008; 46(8):2742-47.
- Wong JY, Chye FY. Antioxidant properties of selected tropical wild edible mushrooms. Journal of Food Composition and Analysis. 2009;22:269–277.
- Agahar MD, Subbulakshmi G. Nutritional value of edible wild mushrooms collected from the Khasi hills of Meghalaya. Food Chemistry. 2005;89:599–603.
- 17. Liu RH. Health benefits of fruits and vegetables are from additive and synergistic combination of phytochemicals. American Journal of Clinical Nutrition. 2003;78:517S–520S.
- Soobrattee MA, Neergheen VS, Luximon-Ramma A, Aruoma OI, Bahorun T. Phenolics as potential antioxidant therapeutic agents: Mechanism and actions. Mutation Research. 2005;579: 200–213.

- Lindequist U, Niedermeyer THJ, Julich W
 D. The pharmacological potential of mushrooms. 2005;2:285–299.
- Elmastas M, Isildak O, Turkukel I, Temur N. Determination of antioxidant activity and antioxidant compounds in wild edible mushrooms. Journal of Food Composition and Analysis. 2007;20:337–345.
- Lo KM, Cheung PCK. Antioxidant activity of extracts from the fruiting bodies of Agrocybe aegerita var. Alba. Food Chemistry. 2005;89:533-539.
- Acharya K, Rai M. Proximate composition, free radical scavenging and NOS activation properties of a wild edible mushroom. Int J Pharm Pharm Sci. 2013;4:67-72.
- Patra S, Patra P, Maity KK, Mandal S, Bhunia SK, Dey B.A heteroglycan from the mycelia of *Pleurotus ostreatus*: Structure determination and study of antioxidant properties. Carbohydr Res. 2013;368:16-21.
- Samanta S, Maity K, Nandi AK, Sen IK, Devi KSP, Mukherjee S. A glucan from an ectomycorrhizal edible mushroom *Tricholoma crassum* (Berk.) Sacc.: Isolation, characterization and biological studies. Carbohydr Res. 2013;367:33-40.
- Nasir MS, Nadeem AA, Nadia S. Effect of Different Methods of Compost Preparationand Lime Concentration on the Yield of *Pleurotus sajor-caju*. International Journal of Agriculture and Biology. 2006;8: 129-131.
- Singh M, Vijay B, Kamal S, Wakchaure G
 C. Mushrooms: Cultivation, Marketing and Consumption. Directorate of Mushroom Research, Indian Council of Agricultural Research (ICAR), Solan (India); 2011.
- 27. Muthangya M, Mshandete M, Amana MJ, Hashim SO, Kivaisi AK. Nutritional and Antioxidant Analysis of Pleurotus HK 37 Grown on *Agave sisalana* saline solid waste. International Journal of Research in Biochemistry and Biophysics. 2014;4(2):5-
- 28. Michael HW, Bultosa G, Pant LM. Nutritional contents of three edible oyster mushrooms grown n two substrates at Hara¬maya, Ethiopia, and sensory properties of boiled mushroom and mushroom sauce. Inter. J. Food Sci. and Techn. 2011;46(4):732–738.

- Muthangya M, Hashim SO, Amana JM, Mshandete AM, Kivaisi AK. Optimization of Pleurotus mushroom cultivation on saline sisal solid waste. World Applied Sciences Journal. 2013;23(9):1146-1150.
- Windhan WR, Mertens DR, Barton li FE. Protocol for NIRS calibration: sample selection and equation development and validation In: Definition of NIRS analysis. Near Infrared Reflectance Spectroscopy (NIRS): Analysis of forage quality. Washington: USDA; 1989.
- Tibuhwa DD, Lyantagaye SL, Mshandete AM. Effect of different post-harvest treatments on nutritive and antioxidant activities of wild edible *Coprinus cinereus* (Schaeff.) S. Gray from Tanzania. Int J Res Biol Sci. 2012a;2:150-156.
- Tibuhwa DD. Antiradical and antioxidant activities of methanolic extracts of indigenous termitarian mushroom from Tanzania. Food Science and Quality Management. 2012b;7:13–23.
- Jaita P, Sourav G, Khandakar ST, Krishnendu A. In vitro free radical scavenging activity of wild edible mushroom, Pleurotus squarrosulus (Mont.) Singer. Indian J. Exp. Biol. 2010;47:1210-1218.
- Nagata M, Yamashita I. Simple method for simultaneous determination of chlorophyll and carotenoids in tomato fruit, Nippon Shokuhin Kogyo Gakkaishi. 1992;39:925-928.
- 35. Plummer TD. An introduction to Practical Biochemistry, 3rd Edition, McGraw-Hill Book Company (UK) Limited. England; 1987.
- 36. Masuda T, Mizuguchi S, Tanaka T, Iritani K, Ali AM. Screening of compounds from the rhizomes of Alpiniaspeciosa. Chem. 2000;48:1479–1484.
- Dinis TCP, Madeira VMC, Almeida LM. Action of phenolic derivates (acetoaminophen, salycilate, and 5aminosalycilate) as inhibitors of membrane lipid peroxidation and as peroxyl radical scavengers. Arch. Biochem. Biophy. 1994; 315:161-169.
- Oyetayo VO, Ariyo OO. Micro and Macronutrient Properties of *Pleurotus* ostreatus (Jacq: Fries) Cultivated on Different Wood Substrates. Jordan Journal of Biological Sciences. 2013;6:223-226.
- 39. Chang ST, Miles PG. Mushrooms: Cultivation, Nutritional Value, Medicinal

- Effect, and Environmental Impact (2nd Ed.). CRC Press. Boca Raton FL; 2004.
- Sales-Campos C, Araujo LM, de Minhoni, MT, De Andrade MCN. Physiochemical analysis and centesimal composition of *Pleurotus ostreatus* mushroom grown in residues from the Amazon. Ciênc. Tecnol. Aliment. Campinas. 2011;31(2): 456-461.
- Kurtzman Jr. RH. A review mushrooms: sources for modern western medicine. Micologia Aplicada International. 2005;17: 21-33.
- 42. Akyüz M, Kirba g S. Nutritive value of wild edible and cultured mushrooms. Turk. J. Biol. 2010;34:97–102.
- 43. Bernaś E, Jaworska G, Lisiewska Z. Edible Mushrooms as a Source of Valuable Nutritive Constituents. Acta Scientiarum Polonorum Technologia Alimentaria. 2006:1: 5-20.
- Mshandete AM, Cuff J. Proximate and nutrient composition of three types of indigenous edible wild mushrooms grown in Tanzania and their utilization prospects. African Journal of Food, Agriculture, Nutrition and Development. 2007:7:2–9.
- Bonatti M, Karnopp P, Soares HM, Furlan SA. Evaluation of *Pleurotus ostreatus* and *Pleurotus sajor caju* nutritional characteristics when cultivated in different lignocellulosic wastes. Food Chem. 2004; 88:425-428.
- 46. Kalac P, Svoboda L. A review of trace element concentrations in edible mushrooms. Food Chemistry. 2000;69: 273-281.
- Afiukwa CA, Oko AO, Afiukwa JN, Ugwu OPC, Ali FU, Ossai EC. Proximate and Mineral Element Compositions of Five Edible Wild Grown Mushroom Species in Abakaliki, Southeast Nigeria Research Journal of Pharmaceutical, Biological and Chemical Sciences. 2013;4(2):1056-1064.
- Ahmed SA. Kadam JA, Mane VP, Patil S S, Baig MMV. Biological efficiency and nutritional contents of Pleurotus Florida (Mont.) Singer cultivated on different agrowastes. Nature and Science. 2009;7:44-48.
- Dimitrios B. Sources of natural phenolic antioxidants. Trends Food Sci Tech. 2006; 17:505-512.
- 50. Banerjee S, Sanjay KR, Chethan S, Malleshi NG. Finger millet

- (*Eleusine coracana*) polyphenols: Investigation of their antioxidant capacity and antimicrobial activity. African Journal of Food Science. 2012;6:362–374.
- 51. Jayakumar T, Thomas PA, Geraldine P. *In-vitro* antioxidant activities of an ethanolic extract of the oyster mushroom, Pleurotus ostreatus. Innov. Food Sci. Emerg. Technol. 2009:10:228–234.
- 52. Gonzalez-Nunez D, Claria J, Rivera F, Poch E. Hypertension. 2001;37:334-338.
- 53. Filippos V, Emmanaouil T, Carl D, Guenter V, Georg K, Nikolas P. Biotechnology Journal. 2007;2(10):1214-1234.
- 54. Ogbonnia S, Adekunle AA, Bosa MK, Enwuru VN. Evaluation of acute and subacute toxicity of Alstonia congensis Engler (Apocynaceae) bark and Xylopia aethiopica (Dunal) A. Rich (Annonaceae) fruits mixtures used in the treatment of diabetes. African Journal of Biotechnology. 2008;7(6):701-705.
- 55. Singla R, Ganguli A, Ghosh M. Antioxidant activities and polyphenolic properties of raw and osmotically dehydrated dried mushroom (Agaricus bisporous) snack food. International Journal of Food Properties. 2010;13:1290–1299.
- 56. Ferreira ICFR, Baptista P, Vilas-Boas M, Barros L. Free radical scavenging capacity and reducing power of wild edible mushrooms from northeast Portugal: Individual cap and stipe activity. Food Chem. 2007;100:1511-1516
- 57. Rao AV, Agarwal S. Role of Antioxidant Lycopene in Cancer and Heart Diseases. J Am CollNutr. 2000;19:563.
- Petre MA, Teodorescu E, Ţuluca C, Bejan, Andronescu A. Biotechnology of mushroom pellets producing by controlled submerged fermentation. Rom. Biotechnol. Lett. 2010;15s: 50-55.
- 59. Barros LP, Baptista LM, Estevinho, Ferreira ICFR. Bioactive properties of the medicinal mushroom *Leucopaxillus giganteus* mycelium obtained in the presence of different nitrogen sources, Food Chem. 2007;105:179-186.
- 60. Filipa SR, Isabel CFRF, Barros L, Martins A. A comparative study of tocopherols composition and antioxidant properties of *in vivo* and *in vitro* ectomycorrhizal fungi. Food Sci. Technol. 2011;44:820–824.

- Abdullah N, Ismail SM, Aminudin N, Shuib AS, Lau BF. Evaluation of selected culinary-medicinal mushrooms for antioxidant and ACE inhibitory activities.
- Evidence-Based Complementary and Alternative Medicine. 2012;1-12.
 62. Halliwell B. Drugs and Aging. 2001;18(9), 685–716.

© 2019 Muthangya et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle3.com/review-history/38006